PERSPECTIVES

Control of Bone Formation by Osteocytes? Lessons from the Rare Skeletal Disorders Sclerosteosis and van Buchem Disease

Rutger L. van Bezooijen,^{1,2} Socrates E. Papapoulos,¹ Neveen A. Hamdy,¹ Peter ten Dijke,² and Clemens W. Löwik¹

¹Department of Endocrinology and Metabolic Diseases and ²Molecular Cell Biology, Leiden University Medical Center, Leiden, The Netherlands

In the adult, bone homeostasis is maintained by a tight balance between the supply, activity, and life span of osteoclasts and osteoblasts within the basic multicellular unit (BMU), a temporary anatomic structure within which bone remodeling occurs. Hormones, cytokines, and mechanical factors are known to control this balance. The exact role of osteocytes, the most abundant cell type in bone, remains unclear. Osteocytes are thought to be mechanosensor cells that control the activity of osteoblasts and osteoclasts within a BMU. Marotti and Martin suggested that osteocytes produce an inhibitory factor that signals to osteoblasts at a bone forming surface, causing them to slow down osteoid formation (1;2), but direct evidence for this is lacking. Recent data obtained from detailed analysis of the two closely related, rare skeletal disorders sclerosteosis and van Buchem disease demonstrated that osteocytes indeed produce a negative regulator of bone formation (3;4).

Patients with sclerosteosis and van Buchem disease have similar phenotypes, and both disorders are characterized by a substantial increase in bone mass (5-7). Sclerosteosis is due to premature termination mutations in the SOST gene on chromosome 17q12-q21 (8;9), whereas a 52 kb homozygous deletion downstream of the SOST gene is associated with van Buchem disease (10;11). The SOST gene encodes a protein, named sclerostin, the expression of which is highly restricted to osteocytes in the adult (3;4). Sclerostin is specifically localized in mature osteocytes in mineralized cortical and cancellous bone (12). In patients with sclerosteosis and van Buchem disease, sclerostin is not present in bone (3;7). Using transgenesis and in vitro transfection studies, a candidate enhancer element that

may drive *SOST* expression in bone, but not during digit development, was identified within the van Buchem deletion (13). Syndactyly is a feature of the phenotype in sclerosteosis. Its absence in van Buchem disease may be explained by the absence of regulatory sequences in the van Buchem deletion that determine *SOST* expression in the zone of digit development.

Increased bone mass in these two rare skeletal diseases is due to increased osteoblast activity, as demonstrated by the following histological features in bone biopsies of affected individuals: predominance of cuboibal, active-appearing osteoblasts, increased double tetracycline label spacing, and increased osteoid that mineralizes normally (3;14;15). Osteoclast numbers appear not to be affected. In vitro studies confirm that sclerostin is a negative regulator of bone formation. Transgenic mice with overexpression of sclerostin have been found to be osteopenic (4;13).

Sclerostin Is Not a Classical BMP Antagonist

Based on amino acid sequence similarity, sclerostin was suggested to be a member of the DAN family of glycoproteins (9). This family of proteins includes wise, cerberus, DAN, coco, caronte, gremlin, dante, and protein related to DAN and cerberus (PRDC) that share as their main characteristic the ability to antagonize bone morphogenetic protein (BMP) activity. Of the BMP antagonists described so far, chordin and noggin have been shown to antagonize BMP signaling by blocking the binding of BMPs to their receptors (16;17). A similar mechanism has been proposed for members of the DAN family.

BMPs are secreted cytokines that were originally identified because of their ability to induce ectopic bone and cartilage formation (18). They exert their effects through distinct combinations of two different types of serine/threonine kinase receptors, *i.e.*, type I and type II receptors (19;20). In initial studies, sclerostin was found to bind BMPs and to antagonize BMP-stimulated alkaline phosphatase activity in osteoblastic cells (3;4;21). However, detailed analysis of the mechanism by which sclerostin antagonizes BMP-stimulated bone formation revealed that the protein is not a classical BMP antagonist (3).

The capacity of sclerostin to inhibit late BMP responses such as bone formation, without antagonizing early BMP responses such as Smad phosphorylation and BMP reporter construct activation, could be achieved via a BMP-induced co-factor that allows sclerostin to function as a BMP antagonist. This hypothetical co-factor may, for example, increase the relative low binding affinity (10⁻⁸ M) of sclerostin to BMPs (4;21), an effect similar to that of twisted gastrulation for the binding of chordin to BMPs (22). However, we recently found that even after a previous BMP stimulation, sclerostin still did not antagonize BMP signaling (23). Available data thus strongly suggest that sclerostin is not a classical BMP antagonist in osteoblastic cells, although an effect on BMPs that have not been tested thus far cannot be excluded.

Sclerostin Is a Wnt Antagonist

As an alternative to antagonizing BMP signaling, sclerostin may inhibit late BMP responses by antagonizing other factors that cooperate with BMPs to stimulate bone formation. Xenopus cerberus, coco, and wise, three DAN family members of which wise has the highest amino acid similarity with sclerostin, have been found to antagonize Wnt activity. Wnts are known to cooperate with BMPs in stimulating bone formation (24-26). Like BMPs, Wnts are secreted cytokines with pivotal roles in a variety of cellular activities, including cell fate determination, proliferation, migration, polarity, and differentiation (27;28).

Recently, sclerostin was found to bind to the first two YWTD-EGF repeats of Wnt coreceptors LRP5 and LRP6 and to antagonize Wnt1- and Wnt3-stimulated activation of a canonical Wnt reporter construct (29:30). Similarly, we showed in a preliminary short report that sclerostin antagonized Wnt reporter construct activation by both BMPs and ligand independent constitutive active BMP receptors in osteoblastic cells, suggesting that sclerostin antagonized Wnt activity (23). However, sclerostin appears not to antagonize Wnt3A-induced β–catenin stabilization in mouse mesenchymal C3H10T1/2 cells (31), which is consistent with our own observations when we used recombinant Wnt3A. However, when we used Wnt expression vectors to stimulate Wnt signaling, sclerostin did antagonize Wnt signaling (23). In all studies reported so far which sclerostin antagonized Wnt in signaling, Wnt expression vectors were used as the source of Wnt activity. The differences between the effects of Wnts produced by transiently transfected cells and recombinant Wnts suggests that these molecules are differentially recognized by sclerostin. This may be explained by differences in tertiary structure, glycosylation, and/or other characteristics of the Wnts. Alternatively, Wnts produced by transient transfection may be membranebound, and this may be important for the interference by sclerostin. The mechanism by which sclerostin binding to LRP5 and 6 antagonizes Wnt signaling is currently unclear, although, for example, Wnt3 did not appear to compete with sclerostin for binding to LRP5 (29).

A Common Signaling Pathway is Affected in Sclerosteosis, van Buchem Disease, and Human High Bone Mass Phenotype

The human high bone mass (HBM) phenotype is an autosomal dominant condition that, like sclerosteosis and van Buchem disease, is characterized by increased bone mass due to enhanced bone formation in the presence of normal bone resorption (32). In two North American Caucasian families with the HBM phenotype, a G/T substitution at position 512 in exon 3, encoding a glycine/valine substitution at amino acid residue 171 (G171V) of the LRP5 gene that makes it resistant to Dkk1-mediated inhibition, was identified as the underlying genetic defect (33-35). The G171V mutation and other mutations in *LRP5* associated with the HBM phenotype were recently shown to have little

effect on LRP5 transit to the cell surface, but instead acted by reducing the affinity for and inhibition by Dkk1, thereby increasing Wnt signaling (36).

Although sclerosteosis, van Buchem disease, and HBM have similar skeletal phenotypes, two distinct molecular mechanisms, increased BMP and Wnt signaling, were believed to cause these disorders. The recent observations that sclerostin antagonizes Wnt signaling rather than BMP signaling raises the possibility that the skeletal disorders sclerosteosis and van Buchem disease, as well as HBM, are due to increased activity of the same signaling pathway: LRP5-mediated canonical Wnt signaling. In the HBM phenotype, the inability of Dkk1 to inhibit LPR5-mediated Wnt signaling increases bone formation, while in sclerosteosis and van Buchem disease, the phenotype is due to the inability of sclerostin to inhibit LPR5-mediated Wnt signaling. LRP5 mutations associated with the HBM phenotype may cause reduced binding and inhibition by sclerostin, in addition to reduced affinity and inhibition by Dkk1.

Conclusion

Evidence obtained during the past 2 years indicates that sclerostin is an osteocyteexpressed protein that inhibits the activity of osteoblasts and prevents them from promoting excessive bone formation. Sclerostin may be transported by the canaliculi to the bone surface, where it inhibits the bone-forming activity of osteoblasts. In this respect, it serves the function of the unknown inhibitory factor, proposed by Marotti and Martin, that is secreted by mature osteocytes and communicates with osteoblasts at a forming surface, causing them to slow osteoid formation (1;2). Alternatively, sclerostin may have an autocrine negative regulatory effect on Wnt signaling in osteocytes and, thereby, inhibit osteocyte-directed indirectly osteoblastic bone formation. Wnt signaling was recently reported in osteocytes using Wnt activity reporter mice (37).

Disturbances in bone remodeling balance constitute the pathophysiological basis of common skeletal disorders such as osteoporosis. It is obvious that inhibition of the activity or production of sclerostin is a promising strategy for the development of therapeutics that stimulate bone formation, thereby increasing bone mass. As sclerostin is a secreted protein, one approach to achieve this is to develop humanized neutralizing monoclonal antibodies capable of inhibiting the biological activity of sclerostin. A preliminary short report indicates that such an approach has been successful in rats (38). There have been concerns, however, about attempts to stimulate bone formation by targeting the sclerostin/LRP5 axis, as such therapeutics may lead to unwanted skeletal side effects. We have recently shown that heterozygous carriers of sclerosteosis have bone mineral density values consistently higher than healthy subjects, without any of the bone complications encountered in homozygotes (39). This suggests that the production and/or activity of sclerostin might be titrated in vivo to promote increases in bone mass without necessarily leading to unwanted skeletal side effects.

The only currently available therapy to stimulate bone formation in humans, intermittent parathyroid hormone (PTH) administration, was recently shown to reduce *SOST* mRNA and sclerostin protein expression in rats and mice (40;41). This suggests that inhibition of sclerostin expression, thereby removing a negative regulator of Wnt-stimulated bone formation, may play a role in the bone formation stimulating effect of intermittent PTH therapy.

Understanding the molecular mechanism of sclerostin action may not only provide the basis for addressing issues in the management of individuals with osteoporosis, but may also help in the management of affected individuals with sclerosteosis or van Buchem disease, for whom the only currently available treatment is surgical removal of excess bone, a difficult and risky procedure because of its anatomical location.

Conflict of Interest: The authors report that no conflicts of interest exist.

BoneKEy-Osteovision. 2005 December;2(12):33-38 http://www.bonekey-ibms.org/cgi/content/full/ibmske;2/12/33 DOI: 10.1138/20050189

References

- 1. Marotti G. The structure of bone tissues and the cellular control of their deposition. *Ital J Anat Embryol.* 1996 Oct-Dec;101(4):25-79.
- 2. Martin RB. Does osteocyte formation cause the nonlinear refilling of osteons? *Bone*. 2000 Jan;26(1):71-8.
- van Bezooijen RL, Roelen BA, Visser A, van der Wee-Pals L, de Wilt E, Karperien M, Hamersma H, Papapoulos SE, ten Dijke P, Löwik CW. Sclerostin is an osteocyte-expressed negative regulator of bone formation, but not a classical BMP antagonist. *J Exp Med.* 2004 Mar 15;199(6):805-14.
- Winkler DG, Sutherland MK, Geoghegan JC, Yu C, Hayes T, Skonier JE, Shpektor D, Jonas M, Kovacevich BR, Staehling-Hampton K, Appleby M, Brunkow ME, Latham JA. Osteocyte control of bone formation via sclerostin, a novel BMP antagonist. *EMBO J.* 2003 Dec 1;22(23):6267-76.
- Beighton P, Barnard A, Hamersma H, van der Wouden A. The syndromic status of sclerosteosis and van Buchem disease. *Clin Genet*. 1984 Feb;25(2):175-81.
- 6. Beighton P. Sclerosteosis. *J Med Genet*. 1988 Mar;25(3):200-3.
- van Bezooijen RL, ten Dijke P, Papapoulos SE, Löwik CW. SOST/sclerostin, an osteocyte-derived negative regulator of bone formation. *Cytokine Growth Factor Rev.* 2005 Jun;16(3):319-27.
- Balemans W, Ebeling M, Patel N, Van Hul E, Olson P, Dioszegi M, Lacza C, Wuyts W, Van Den Ende J, Willems P, Paes-Alves AF, Hill S, Bueno M, Ramos FJ, Tacconi P, Dikkers FG, Stratakis C, Lindpaintner K, Vickery B, Foernzler D, Van Hul W. Increased bone density in sclerosteosis is due to the deficiency of a novel secreted protein (SOST). *Hum Mol Genet*. 2001 Mar 1;10(5):537-43.
- Brunkow ME, Gardner JC, Van Ness J, Paeper BW, Kovacevich BR, Proll S, Skonier JE, Zhao L, Sabo PJ, Fu Y,

Alisch RS, Gillett L, Colbert T, Tacconi P, Galas D, Hamersma H, Beighton P, Mulligan J. Bone dysplasia sclerosteosis results from loss of the SOST gene product, a novel cystine knot-containing protein. *Am J Hum Genet*. 2001 Mar;68(3):577-89.

- Balemans W, Patel N, Ebeling M, Van Hul E, Wuyts W, Lacza C, Dioszegi M, Dikkers FG, Hildering P, Willems PJ, Verheij JB, Lindpaintner K, Vickery B, Foernzler D, Van Hul W. Identification of a 52 kb deletion downstream of the SOST gene in patients with van Buchem disease. J Med Genet. 2002 Feb;39(2):91-7.
- Staehling-Hampton K, Proll S, Paeper BW, Zhao L, Charmley P, Brown A, Gardner JC, Galas D, Schatzman RC, Beighton P, Papapoulos S, Hamersma H, Brunkow ME. A 52-kb deletion in the SOST-MEOX1 intergenic region on 17q12-q21 is associated with van Buchem disease in the Dutch population. *Am J Med Genet*. 2002 Jun 15;110(2):144-52.
- Poole KE, van Bezooijen RL, Loveridge N, Hamersma H, Papapoulos SE, Löwik CW, Reeve J. Sclerostin is a delayed secreted product of osteocytes that inhibits bone formation. *FASEB J.* 2005 Nov;19(13):1842-4.
- Loots GG, Kneissel M, Keller H, Baptist M, Chang J, Collette NM, Ovcharenko D, Plajzer-Frick I, Rubin EM. Genomic deletion of a long-range bone enhancer misregulates sclerostin in Van Buchem disease. *Genome Res.* 2005 Jul;15(7):928-35.
- Stein SA, Witkop C, Hill S, Fallon MD, Viernstein L, Gucer G, McKeever P, Long D, Altman J, Miller NR, Teitelbaum SL, Schlesinger S. Sclerosteosis: neurogenetic and pathophysiologic analysis of an American kinship. *Neurology.* 1983 Mar;33(3):267-77.
- 15. Hill SC, Stein SA, Dwyer A, Altman J, Dorwart R, Doppman J. Cranial CT findings in sclerosteosis. *AJNR Am J Neuroradiol.* 1986 May-Jun;7(3):505-11.
- Piccolo S, Sasai Y, Lu B, De Robertis EM. Dorsoventral patterning in Xenopus: inhibition of ventral signals by

direct binding of chordin to BMP-4. *Cell*. 1996 Aug;86(4):589-98.

- Zimmerman LB, De Jesus-Escobar JM, Harland RM. The Spemann organizer signal noggin binds and inactivates bone morphogenetic protein 4. *Cell*. 1996 Aug 23;86(4):599-606.
- 18. Urist MR. Bone: formation by autoinduction. *Science*. 1965 Nov 12;150(698):893-9.
- 19. Canalis E, Economides AN, Gazzerro E. Bone morphogenetic proteins, their antagonists, and the skeleton. *Endocr Rev.* 2003 Apr;24(2):218-35.
- 20. van Bezooijen RL, Heldin CH, ten Dijke P. Bone morphogenetic proteins and their receptors. *Nature Encyclopedia of Life Science*. 2005, in press.
- Kusu N, Laurikkala J, Imanishi M, Usui H, Konishi M, Miyake A, Thesleff I, Itoh N. Sclerostin is a novel secreted osteoclast-derived bone morphogenetic protein antagonist with unique ligand specificity. *J Biol Chem.* 2003 Jun 27;278(26):24113-7.
- 22. Chang C, Holtzman DA, Chau S, Chickering T, Woolf EA, Holmgren LM, Bodorova J, Gearing DP, Holmes WE, Brivanlou AH. Twisted gastrulation can function as a BMP antagonist. *Nature*. 2001 Mar 22;410(6827):483-7.
- van Bezooijen RL, Visser A, van der horst G, Karperien M, Papapoulos SE, ten Dijke P, Löwik CWGM. Sclerostin inhibits BMP-stimulated bone formation by antagonizing Wnt signaling. *J Bone Miner Res.* 2005 Sep;20(Suppl 1):S9 [Abstract]
- 24. Bell E, Munoz-Sanjuan I, Altmann CR, Vonica A, Brivanlou AH. Cell fate specification and competence by Coco, a maternal BMP, TGFbeta and Wnt inhibitor. *Development*. 2003 Apr;130(7):1381-9.
- 25. Itasaki N, Jones CM, Mercurio S, Rowe A, Domingos PM, Smith JC, Krumlauf R. Wise, a context-dependent activator and inhibitor of Wnt signalling. *Development*. 2003 Sep;130(18):4295-305.

- 26. Piccolo S, Agius E, Leyns L. Bhattacharyya Grunz S, Η, Bouwmeester T, De Robertis EM. The head inducer Cerberus is а multifunctional antagonist of Nodal, BMP and Wnt signals. Nature. 1999 Feb 25;397(6721):707-10.
- 27. Bejsovec A. Wnt pathway activation: new relations and locations. *Cell.* 2005 Jan 14;120(1):11-4.
- 28. Logan CY, Nusse R. The Wnt signaling pathway in development and disease. *Annu Rev Cell Dev Biol.* 2004;20:781-810.
- 29. Li X, Zhang Y, Kang H, Liu W, Liu P, Zhang J, Harris SE, Wu D. Sclerostin binds to LRP5/6 and antagonizes canonical Wnt signaling. *J Biol Chem*. 2005 May 20;280(20):19883-7.
- 30. Semenov M, Tamai K, He X. SOST is a ligand for LRP5/LRP6 and a Wnt signaling inhibitor. *J Biol Chem.* 2005 Jul 22;280(29):26770-5.
- Winkler DG, Sutherland MS, Ojala E, Turcott E, Geoghegan JC, Shpektor D, Skonier JE, Yu C, Latham JA. Sclerostin inhibition of Wnt-3a-induced C3H10T1/2 cell differentiation is indirect and mediated by bone morphogenetic proteins. *J Biol Chem.* 2005 Jan 28;280(4):2498-502.
- 32. Johnson ML, Gong G, Kimberling W, Recker SM, Kimmel DB, Recker RB. Linkage of a gene causing high bone mass to human chromosome 11 (11q12-13). *Am J Hum Genet.* 1997 Jun;60(6):1326-32.
- Boyden LM, Mao J, Belsky J, Mitzner L, Farhi A, Mitnick MA, Wu D, Insogna K, Lifton RP. High bone density due to a mutation in LDL-receptor-related protein 5. *N Engl J Med.* 2002 May 16;346(20):1513-21.
- Little RD, Carulli JP, Del Mastro RG, Dupuis J, Osborne M, Folz C, Manning SP, Swain PM, Zhao SC, Eustace B, Lappe MM, Spitzer L, Zweier S, Braunschweiger K, Benchekroun Y, Hu X, Adair R, Chee L, FitzGerald MG, Tulig C, Caruso A, Tzellas N, Bawa A, Franklin B, McGuire S, Nogues X, Gong

BoneKEy-Osteovision. 2005 December;2(12):33-38 http://www.bonekey-ibms.org/cgi/content/full/ibmske;2/12/33 DOI: 10.1138/20050189

G, Allen KM, Anisowicz A, Morales AJ, Lomedico PT, Recker SM, Van Eerdewegh P, Recker RR, Johnson ML. A mutation in the LDL receptor-related protein 5 gene results in the autosomal dominant high-bone-mass trait. *Am J Hum Genet*. 2002 Jan;70(1):11-9.

- 35. Zhang Y, Wang Y, Li X, Zhang J, Mao J, Li Z, Zheng J, Li L, Harris S, Wu D. The LRP5 high-bone-mass G171V mutation disrupts LRP5 interaction with Mesd. *Mol Cell Biol.* 2004 Jun;24(11):4677-84.
- 36. Ai M, Holmen SL, Van Hul W, Williams BO, Warman ML. Reduced affinity to and inhibition by DKK1 form a common mechanism by which high bone massassociated missense mutations in LRP5 affect canonical Wnt signaling. *Mol Cell Biol.* 2005 Jun;25(12):4946-55.
- 37. Hens JR, Wilson KM, Dann P, Chen X, Horowitz MC, Wysolmerski JJ. TOPGAL mice show that the canonical Wnt signaling pathway is active during bone development and growth and is activated by mechanical loading in vitro. *J Bone Miner Res.* 2005 Jul;20(7):1103-13.
- Warmington K, Ominsky M, Bolon B, Cattley R, Stephens P, Lawson A, Lightwood D, Perkins V, Kirby H, Moore A, Robinson M, Kostenuik PJ, Simonet WS, Lacey DL, Paszty C. Sclerostin monoclonal antibody treatment of osteoporotic rats completely reverses one year of overiectomy-induced systemic bone loss. J Bone Miner Res. 2005 Sep;20(Suppl 1):S22 [Abstract]
- Gardner JC, van Bezooijen RL, Mervis B, Hamdy NA, Löwik CW, Hamersma H, Beighton P, Papapoulos SE. Bone mineral density in sclerosteosis; affected individuals and gene carriers. *J Clin Endocrinol Metab*. 2005 Dec;90(12):6392-5.
- 40. Keller H, Kneissel M. SOST is a target gene for PTH in bone. *Bone*. 2005 Aug;37(2):148-58.
- 41. Bellido T, Ali AA, Gubrij I, Plotkin LI, Fu Q, O'Brien CA, Manolagas SC, Jilka RL. Chronic elevation of parathyroid hormone in mice reduces expression of sclerostin by osteocytes: a novel

mechanism for hormonal control of osteoblastogenesis. *Endocrinology*. 2005 Nov;146(11):4577-83.